

Brettanomyces Research Findings and Management

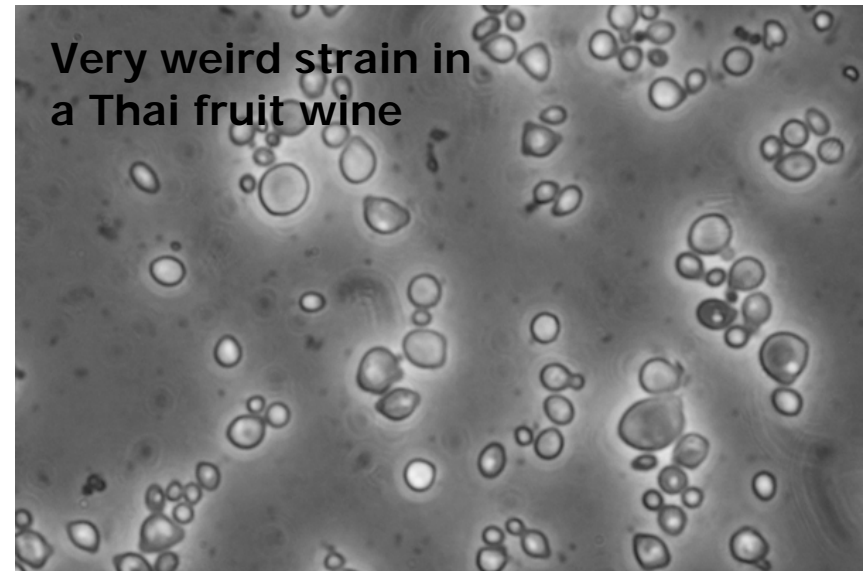
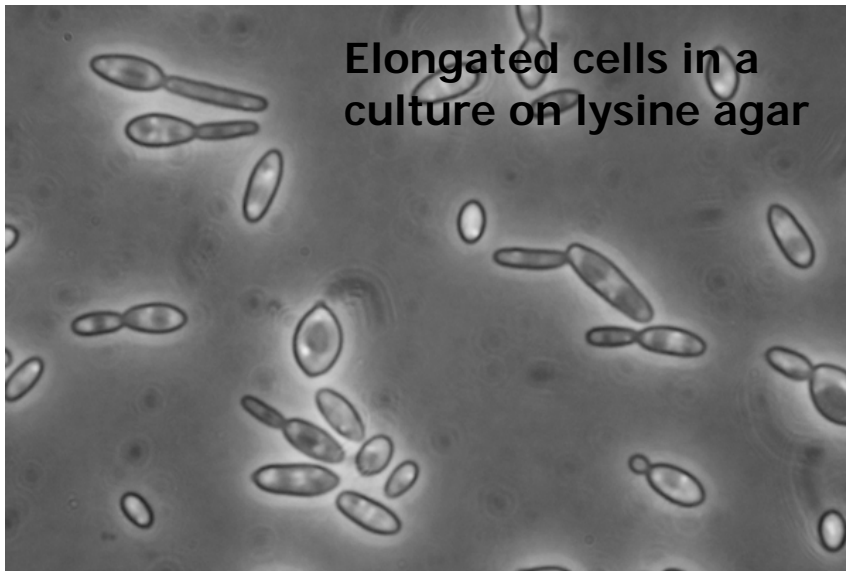
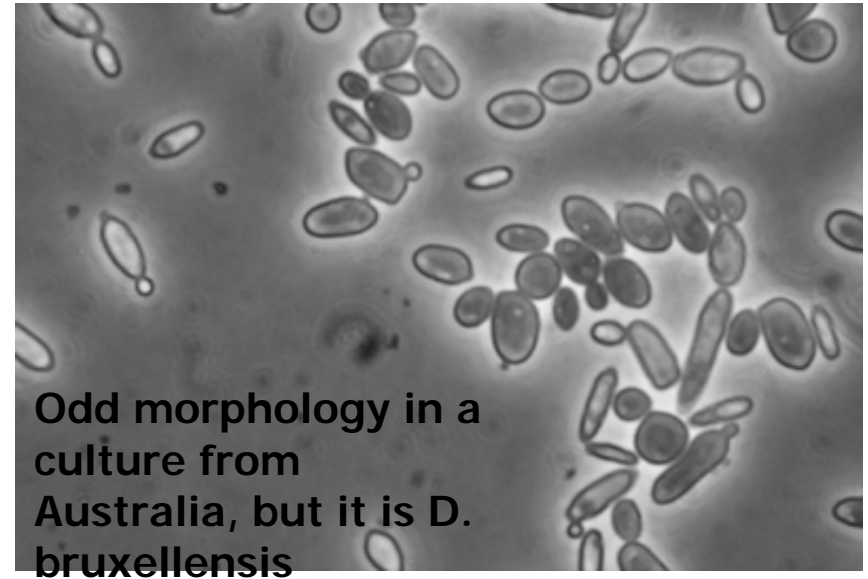
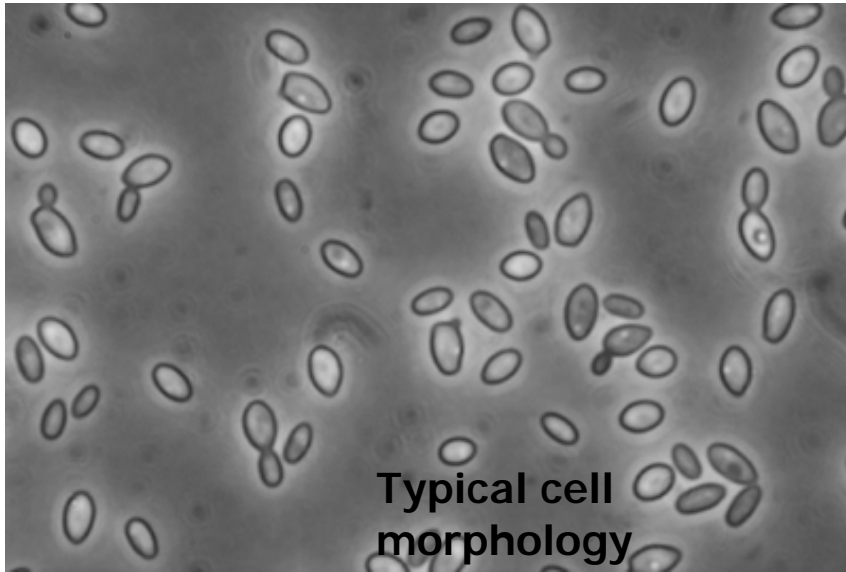
Wine/Enology-Grape Chemistry Group



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www.vtwines.info
Enology Notes #92

The many faces of *Dekkera/Brettanomyces*...



Source: Lisa Van de Water

Brett Descriptors

- **Positive**
 - **Complex**
 - **Mature**
 - **Spicy**
- **Negative (partial list)**
 - **Animals**
 - **Sweaty horse/saddle**
 - **Wet dog**
 - **Manure**
 - **Barnyard**
 - **Mousy aftertaste**
 - **Plastic**
 - **Band-aids**
 - **Burnt plastic**
 - **Other**
 - **Burnt beans**
 - **Rancid**
 - **Metallic**

**Population dynamics and effects of
Brettanomyces bruxellensis strains on
Pinot noir wines**

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***Am. J. Enol. Vitic.* 54:294-300**

Brettanomyces bruxellensis:
**Comparison of Growth Profiles and Metabolites
among Ten Strains in Pinot Noir Wine**

- ***Question: Can differences in winemaker's experiences with Brettanomyces growth in wine be attributed to strain differences?***

Experimental Design:

Ten genetically-characterized strains of *B. bruxellensis*

- **Pinot noir: 30 mg/L sulfur dioxide at crush. Ferment to dryness, press, clarify at 5°C (6 weeks).**
- **Rack to sterile containers, DMDC @ 700 mg/L.**
- **Bottle.**
- **Initial inoculum: 50 CFU/mL (10 strains x 4 replications) + controls.**



Sampling

Weekly samples were plated for growth and chemical analysis for up to 712 days.

Analyte quantification by HE-SPME, GC/MS:

4-Ethylphenol (4-EP)

4-Ethylguaiacol (4-EG)

2-phenylethanol

Guaiacol

Isovaleric acid

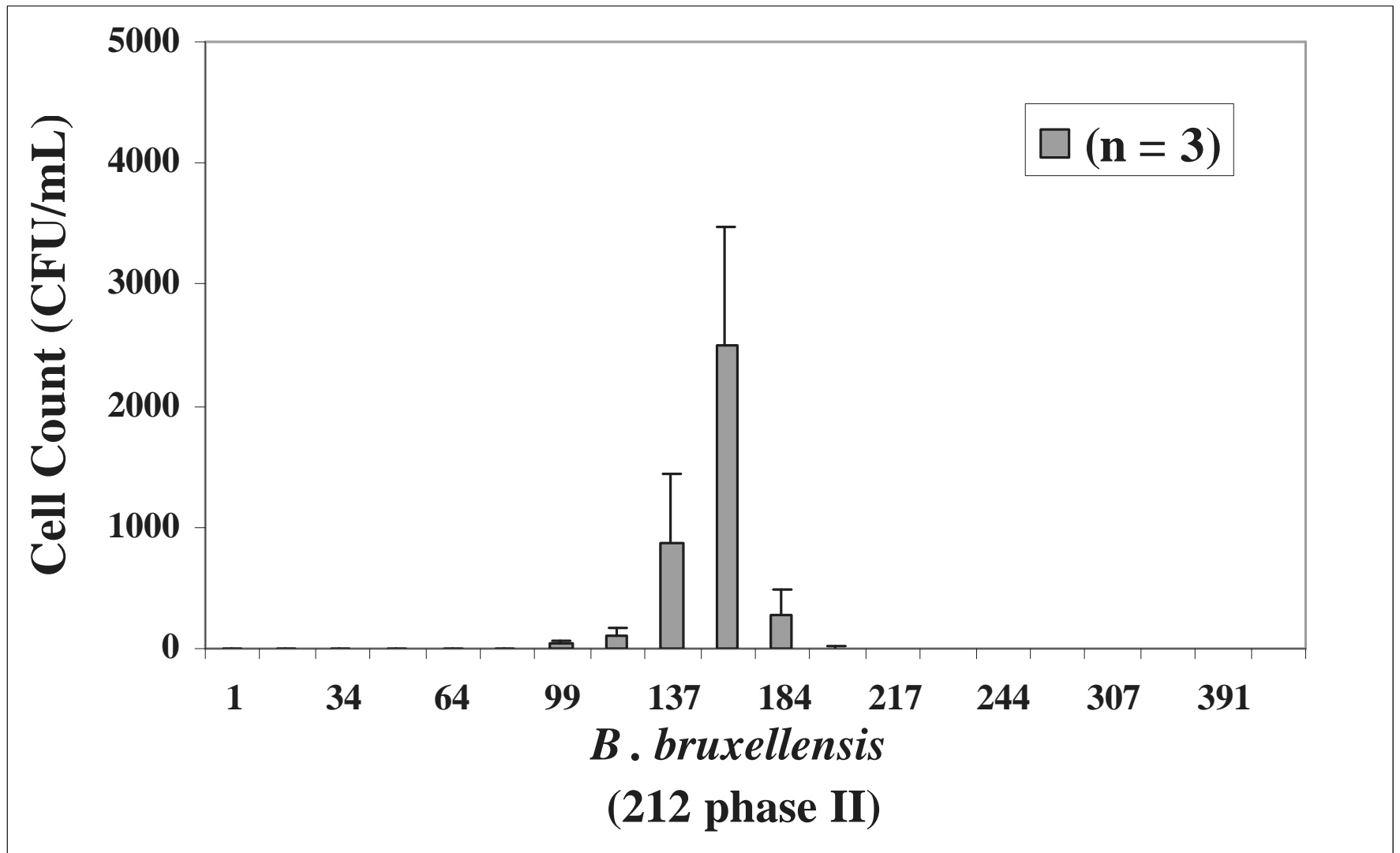
Ethyldecanoate

trans-2-Nonenal

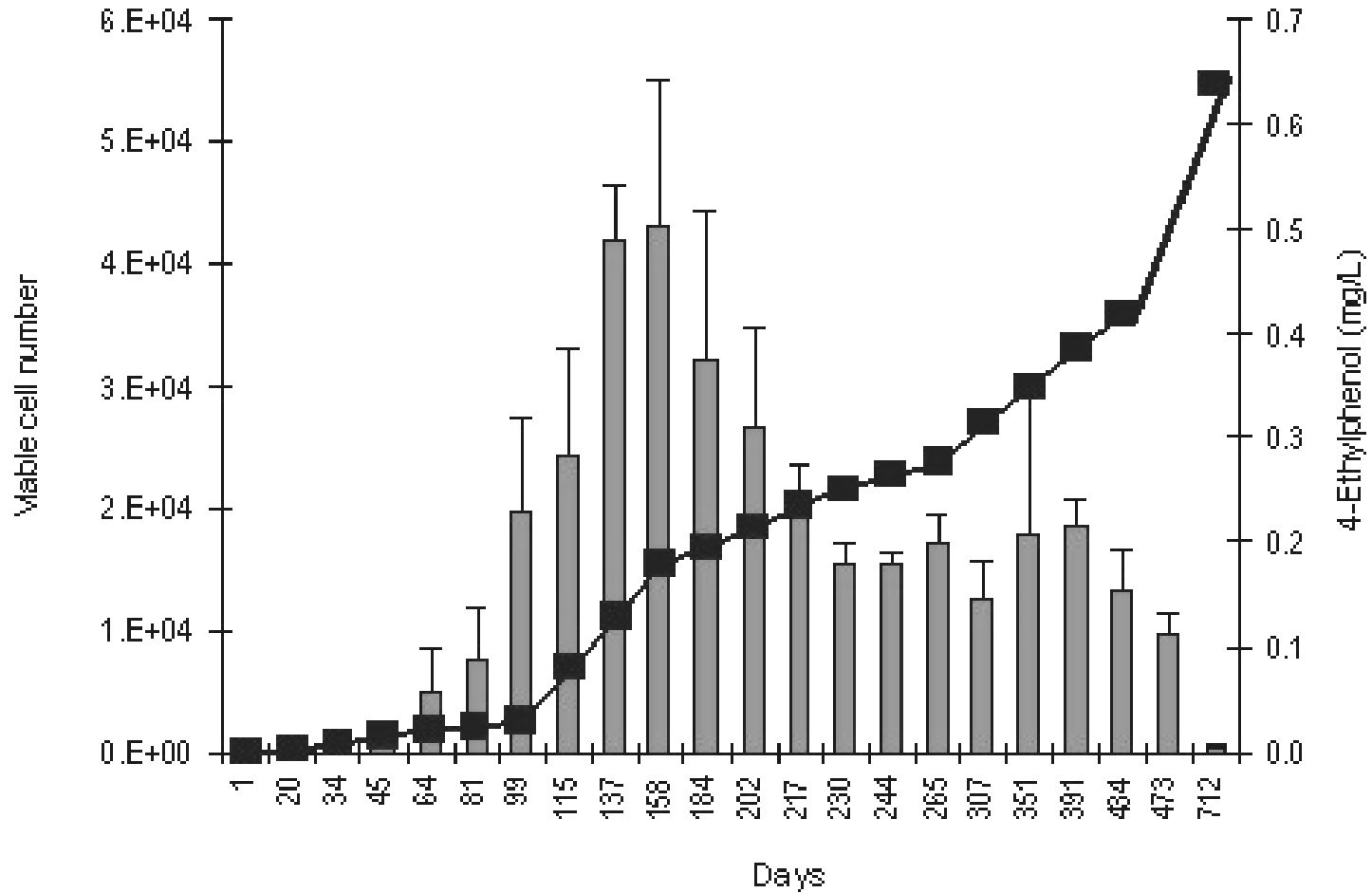
Isoamyl alcohol

Ethyl-2-methylbutyrate

Results

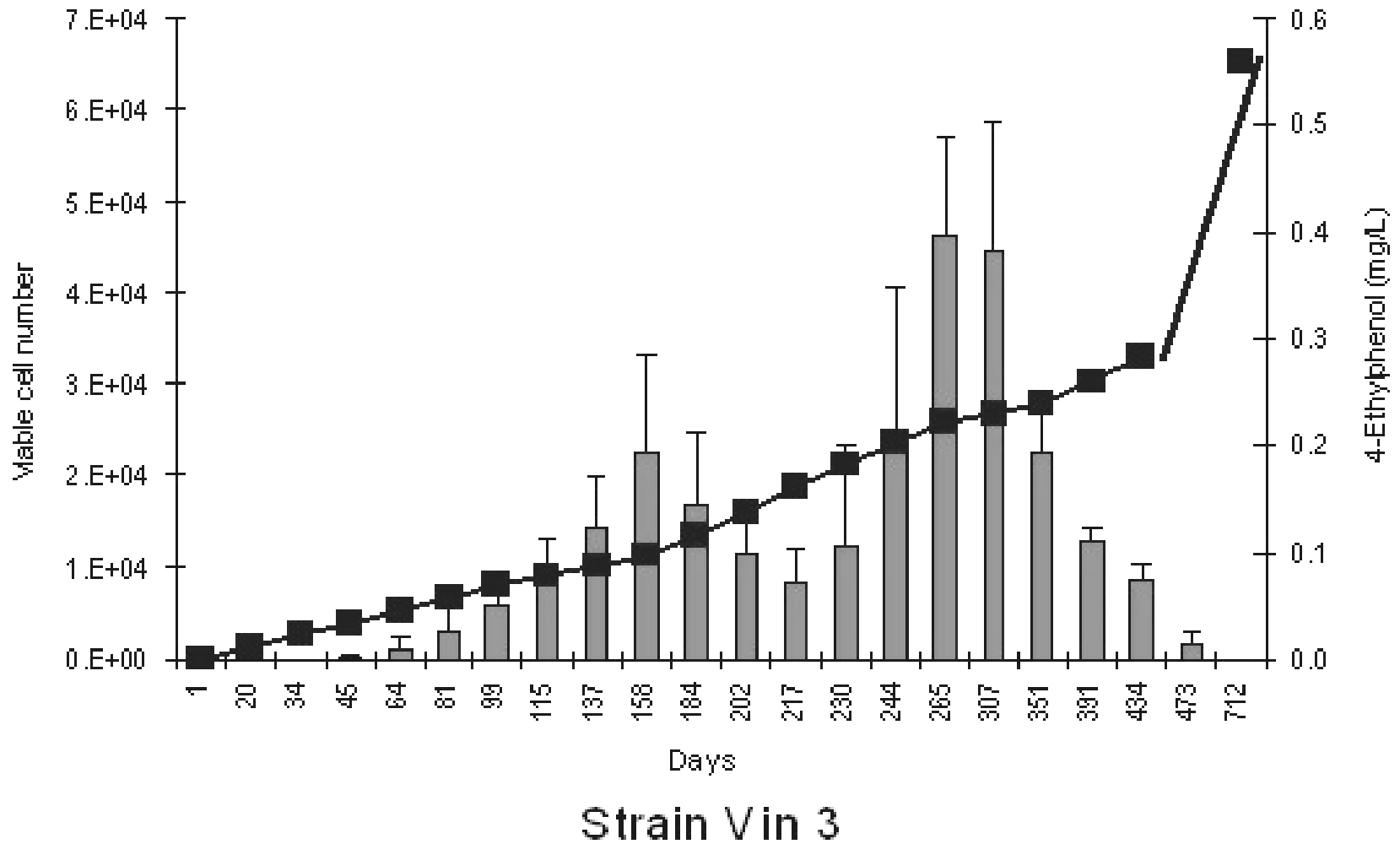


Results (cont.)

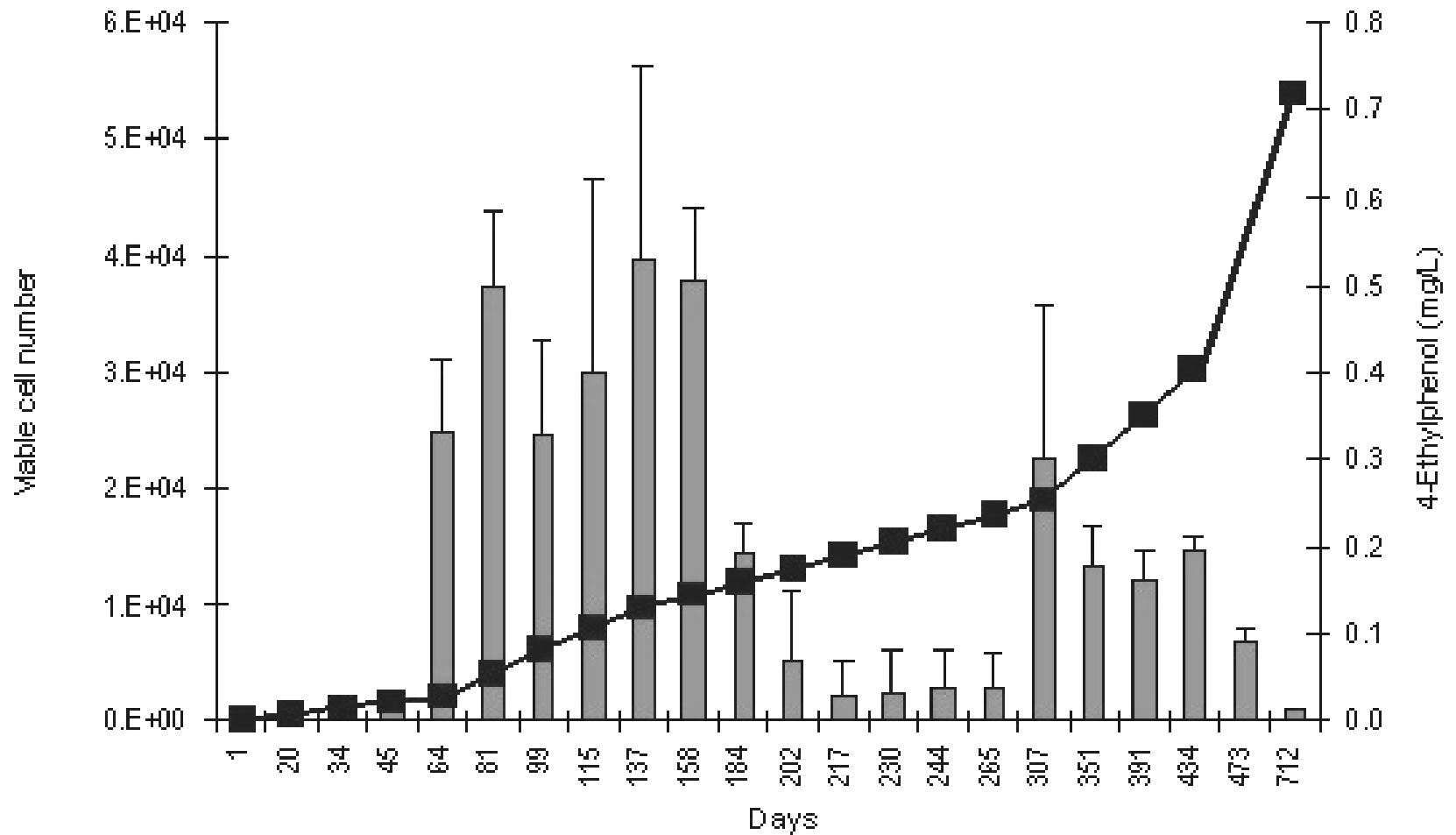


Strain 213

Results (cont.)



Results (cont.)

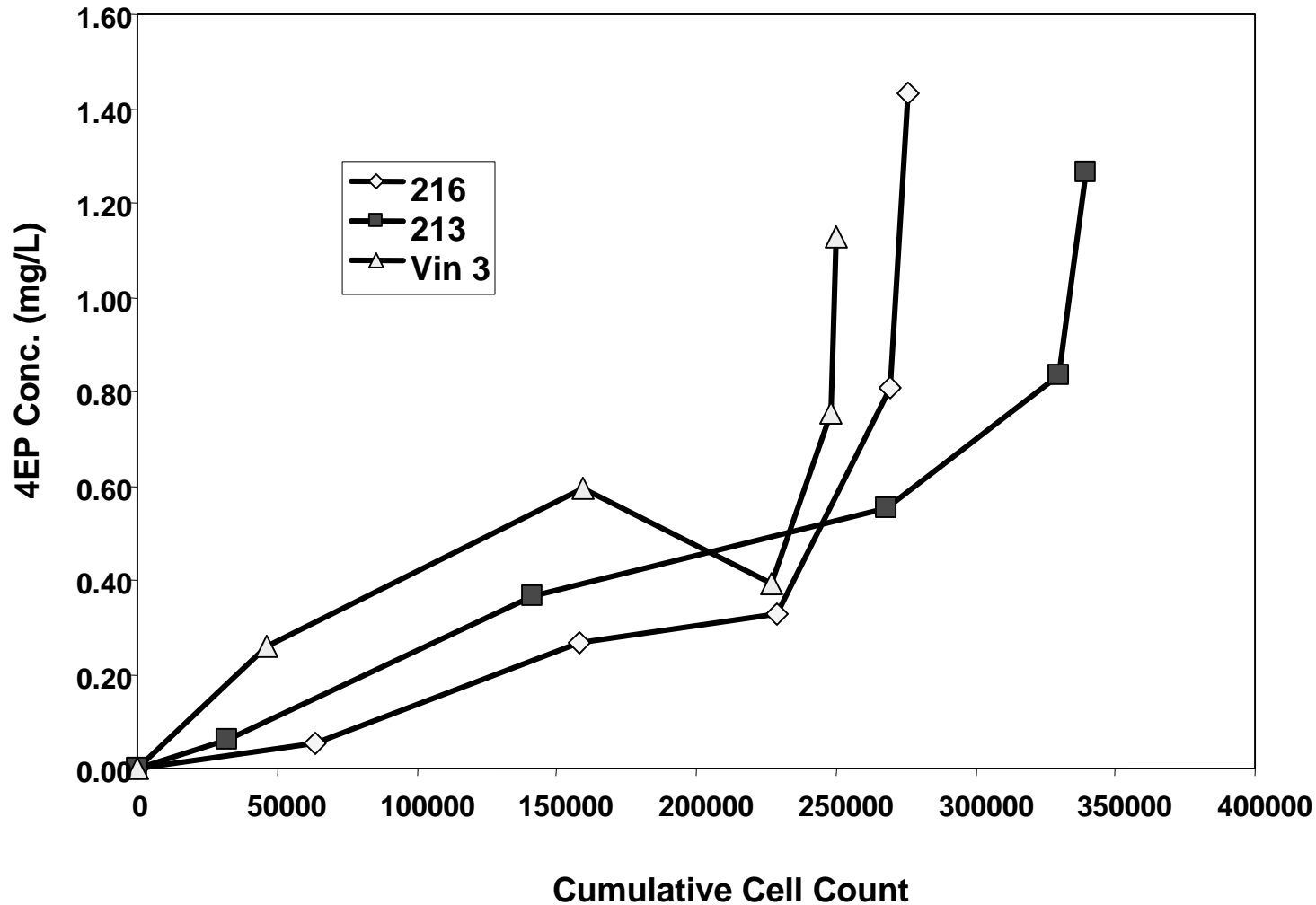


Strain 216



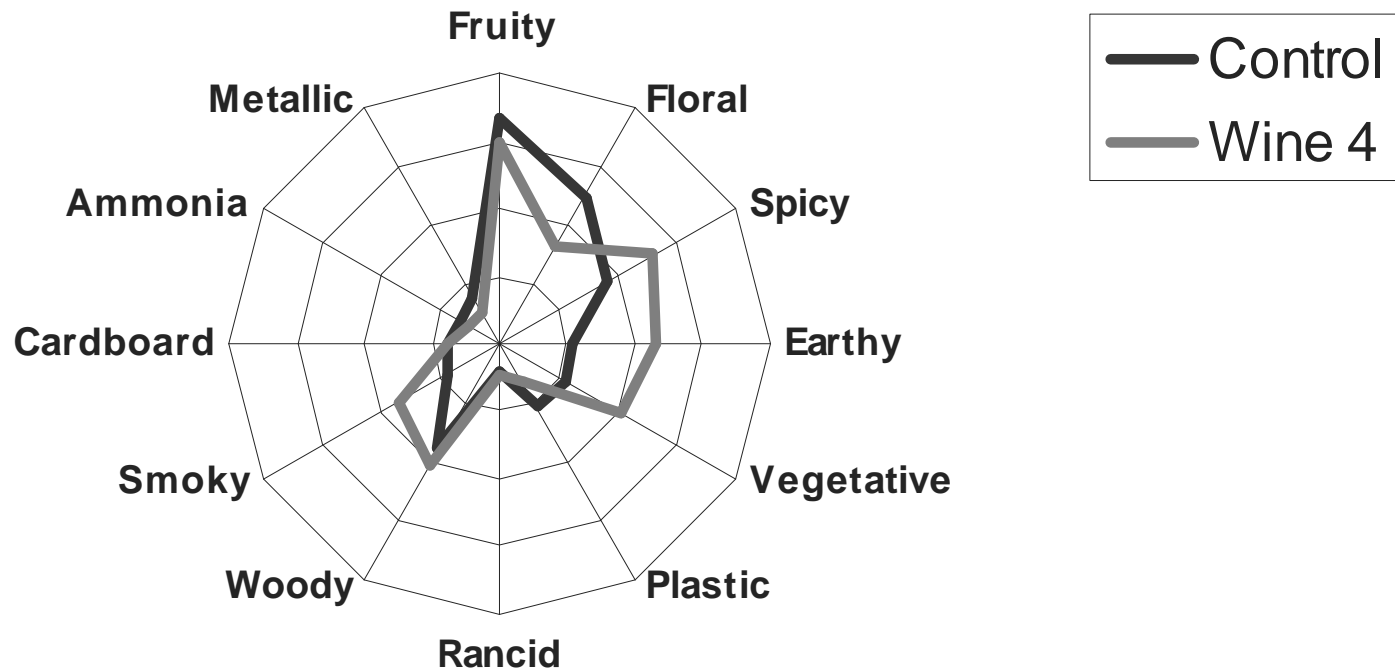
Brettanomyces

4-EP vs Cum. Cell Count, averaged



Sensory Evaluation

Brettanomyces Sensory





Conclusions

- **Significant strain differences in length of growth cycle and peak population densities.**
- **Blooms may be explained by VNC.**
- **Large range of 4-ethylphenol (4-EP).**
- **Large range of 4-ethylguaicol (4-EG).**
- **4-EP and 4-EG correlated.**
- **4-EP and 4-EG not correlated to isovaleric acid (IVA).**

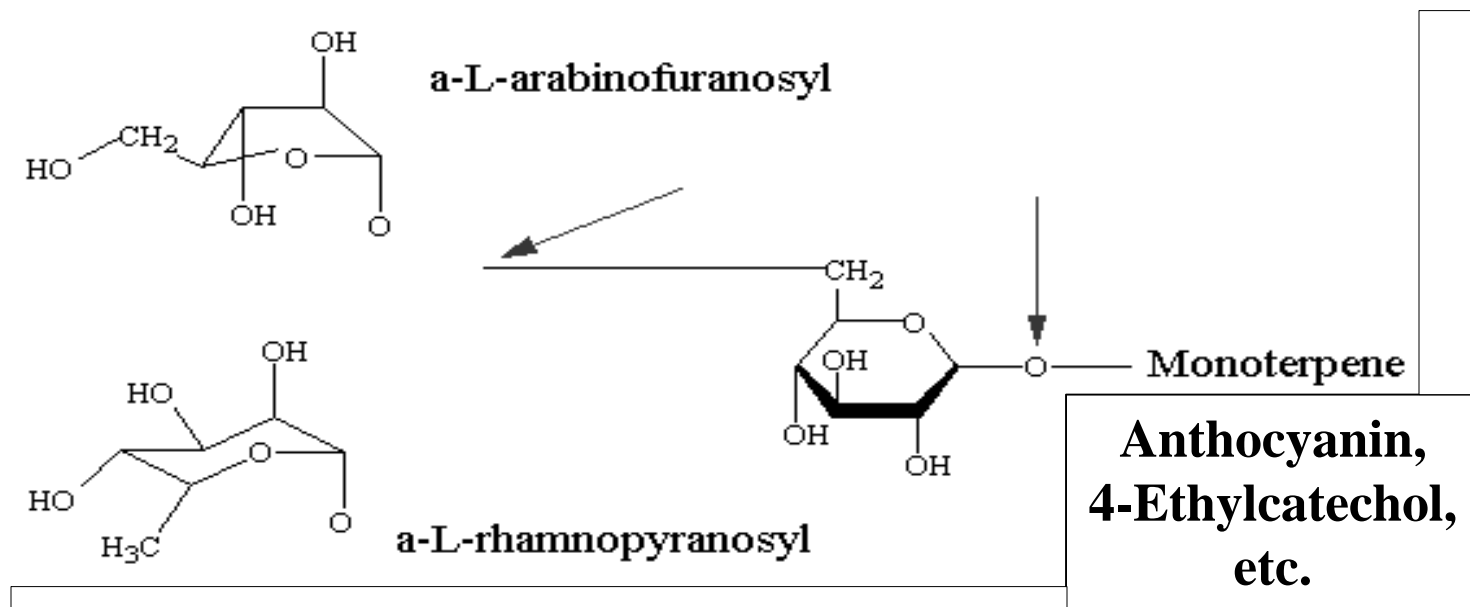


Conclusions

- **With the exception of one strain, most 4-EP was produced after the population reached maximum cell density.**
- **The correlation between 4-EP and viable cell density was not as strong as the correlation with cumulative cell density.**
- **There were significant sensory differences among strains.**
- **4-EP correlated to low glucose/fructose.**

Important Enzymes: Esterases, Glucosidases

- **Glycosidases**



**Glycosidase Activity in *Brettanomyces
bruxellensis*, other Yeasts,
and *Oenococcus oeni***

**H.M. McMahon and B.W. Zoecklein.
J. Ind. Micro. Biotech. 23:198-203.**

**A.K. Mansfield and B.W. Zoecklein.
Am. J. Enol. Vitic. 53:303-307.**

<i>B. bruxellensis</i> strain	Whole Cell	Permeabilized	Supernatant
211	27 c	142 e	11 bcd
212	5 d	341 a	9 bcd
213	34 c	105 f	14 bcd
214	19 c	110 f	6 cd
215	< LOD	74 g	11 bcd
216	59 b	321 b	24 a
Brux	26 c	182 d	11 bcd
Souche 'Ave'	14 c	138 e	7 cd
Souche 'O'	< LOD	< LOD	4 d
Souche 'M'	82 a	179 d	9 bcd
Vin 1	32 c	14 g	< LOD
Vin 3	22 c	232 c	9 bcd
Vin 4	65 b	25 g	4 d
Vin 5	4 d	21 g	11 bc



Conclusions

- **Large variation among strains in total enzyme activity.**
- **Eight strains of *Brettanomyces bruxellensis* had high *beta*-glucosidase activity (670-2,650 nM/mL/g dry cells).**
- **Large variation in supernatant and permeabilized activity.**

Results of Physiological Tests

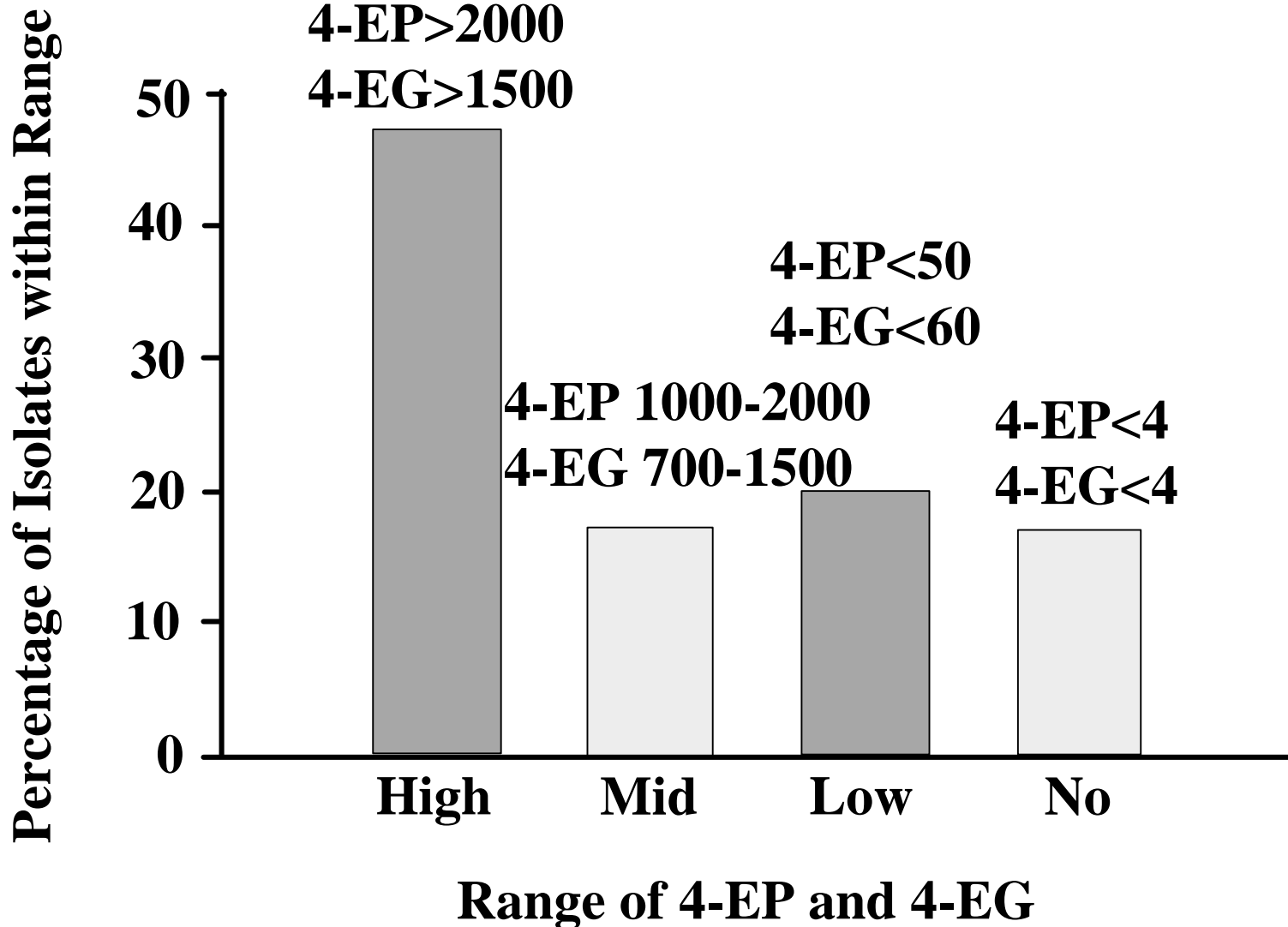
L. Joseph, T. Henick-Kling, L. Conterno

- **Regional differences in metabolism**
 - **75% of European strains used malic acid, 12% CA strains did**
 - **All CA strains used nitrate, < 30% of European strains did**
 - **63% of European strains used ethanol, 18% CA strains did**
 - **Most CA strains grew at 37 C, no European strains did**

Physical Characteristics

- **All isolates tolerant to 10% ethanol or higher.**
- **33 isolates grew well at pH 2.**
- **More than 30% of isolates grew at 10° C.**
- **More than 35% of isolates grew at 37° C.**
- **3 isolates (about 10%) grew at both temperature extremes.**
- **Almost 50% showed tolerance to 30 mg/L or greater free SO₂ at pH 3.4.**

4-EP and 4-EG Production



Climate Impact on *Brett* Metabolites Henschke, 2004

- **4-EP / 4-EG decrease in cool regions**
- **Malvidin-3-*p*-coumaryl glucoside may be precursor to 4-EP**

Brett Growth

- **Physical effects**
 - Usually grows slowly, over many months
 - Can grow within weeks if conditions are favorable
 - Grows in the wine, almost never as a surface film
 - Growth is stimulated by oxygen, but very little is required
 - Slight CO₂ gas
 - Sediment in bottle
- **Sensory effects**
 - Reduced varietal character
 - Esterase activity degrades some fruity aromas
 - Floral aromas are also reduced
 - Aromatic compounds
 - Bitter/metallic finish
 - Sometimes: mousy taint (ACPY/ACTPY)
- **Lots of strain variation**

Brettanomyces Detection

- **Sensory**
 - Train lab and production crew to recognize danger signals using standards
 - When sensory effects are noticeable, it may be too late
- **Matrix effect**

Brett Standards

- **Components of FlavorSense, San Rafael, CA,
Brett standard**
 - **4-Ethylphenol**
 - **4-Ethylguaiacol**
 - **Furfural**
 - **3-methyl-2-buten-1-ol**
 - **Guaiacol**
 - **Isobutyl alcohol**
 - **Isobutyric acid**
 - **Isovaleric acid**
 - **Propionic acid**

***Brettanomyces* Detection**

- **Direct Microscopic Examination**
 - **Difficult when < 1000 cells/ml**
 - **Requires skill in identifying cells**
- **Culturing**
 - **Sampling method is very important**
 - **Detects only microbes that are present and alive**
 - **Disadvantages:**
 - **Must select and prepare media properly**
 - **False negatives (VNC)**
 - **Takes time for growth (3-7+ days)**
 - **Requires skill in identifying colonies**

Brettanomyces Detection

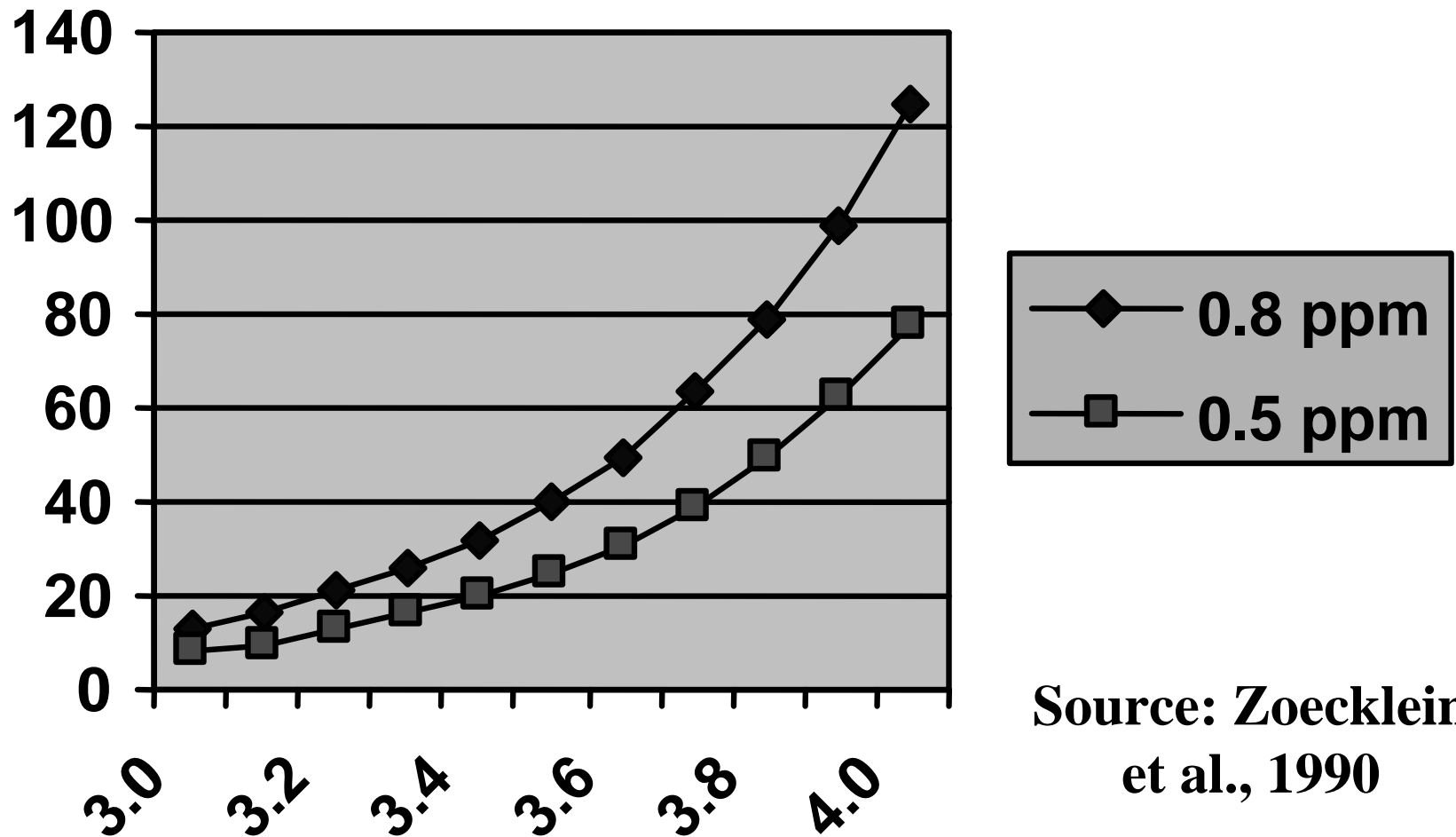
- **Chemical analysis**
 - ‘Marker’ compounds: 4-EP, 4-EG, fatty acids
 - Tests metabolites, not activity itself
 - Not necessarily directly related (e.g., 4-EP vs. culturing)
- **Antibody-based methods**
 - **ELISA**
 - A. Kuniyuki et al. *Am. J. Enol. Vitic.* 35:143-145
- **Nucleic Acid-based methods**
 - **Polymerase Chain Reaction (PCR)**
 - J. Ibeas et al. *Appl. Environ. Microbiol.* 62:998-1003
 - L. Cocolin et al. *Appl. Environ. Microbiol.* 70: 1347-1355
 - T. Phister and D. Mills *Appl. Environ. Microbiol.* 69:7430-7434
- **Problems: False positives, expensive, hand-held ‘field’ unit**

Keys to *Brettanomyces* Management

Generally find only 1 biotype in a wine

- **Wine composition**
 - **Minimize substrates for growth**
 - N (Formol titration, www.vtwines.info or *Am. J. Enol. Vitic.* 53:325-329.)
 - **SO₂ management**
 - **pH management**
- **Winemaking operations**
 - **Cellar temperature**
 - **Population monitoring and control**
 - **Cellar and barrel sanitation / hygiene**
 - **Preparation for bottling**

Free SO₂ Needed to Achieve 0.5 and 0.8 ppm Molecular SO₂, at Different pHs



Source: Zoecklein
et al., 1990

Encourage/Discourage *Brett*

- **To ENCOURAGE**

- Wine composition**

- **Red wine**
 - **pH > 3.6**
 - **Molecular SO₂ < 0.2 mg/L**
 - **Alcohol 13% or below**
 - **Residual hexose sugars**
 - **Biotin, thiamine**
 - **Amino acids**
 - **Yeast lees present**

- **To DISCOURAGE**

- Wine composition**

- **White wine**
 - **pH < 3.6**
 - **Molecular SO₂ 0.4 mg/L or greater**
 - **Alcohol > 13%**
 - **RS < 0.2 g/L**
 - **Vitamins depleted**
 - **“Nutrient desert”**
 - **Clarified**

Encourage/Discourage *Brett*

- **To ENCOURAGE**
Winemaking operations

- Temperature 25-30 C
- Oxidative conditions
- New barrels
- Poor sanitation
- Cross-contamination
- Barrels washed in cold water
- No aggressive barrel sanitation

- **To DISCOURAGE**
Winemaking operations

- Temperature < 16 C
- Keep containers topped / closed
- Older but uninfected barrels
- Good hygiene
- Keep infected wine separate
- High-pressure hot water wash
- Ozone/burn sulfur wick in barrel

Effect of Barrique Sanitation Procedures -

Manuel Malfeito-Ferreira, 2004

- **Barrel sanitation experiment**
 - Cold rinse, then hot water rinse 3x 70 C
 - Same as above plus SO₂ 1 month (200 ppm pH3)
 - Cold rinse, fill with 90 C water 15 min
 - Cold rinse, 70 C rinse, steam low pressure 10 min
 - Most effective treatment
- *Brett / Dekkera* was found 8 mm deep in staves.

Barrels cannot be “sterilized” with SO₂, rinsing, or ozone.

Isolate *Brett*+ barrels.

Ozone Treatment

- **High-pressure water wash barrel**
 - Thorough blast with sharp stream of hot water
 - Rinse for 2-3 minutes
 - Must remove all organics
 - Cool down completely
- **Treat with ozonated water**
 - Filter and deionize water before ozonating
 - At least 2-2.5 mg/L ozone in barrel, 0.1 mg/L out
 - Time x Concentration

***Brett* and Biofilms**

- **Liquid / solid interface**
- **17 / 35 strains form biofilms (Joseph, 2004)**
- **pH effect**
- **Impact of cleaning compounds on biofilms**



Monitoring *Brett*

- **Have a HACCP-like plan (www.vtwine.info)**
- **Isolate contaminated barrels**
- **Sample barrels with disposable plastic pipets**
- **Top with *Brett*-free wine (filtered, pasteurized and/or Velcorin-DMDC)**
- **Keep barrels topped-up or not opened**
- **Monitor carefully before bottling**

Acknowledgements

- **California Agricultural Technology Institute**
- **American Vineyard Foundation**
- **Lallemant, Inc., Montreal, Québec, Canada**
- **Napa Valley Wine Technical Group**
- **Virginia Grape Growers Association**
- **Vino Farms**
- **Roy Thornton**
- **Lisa Van de Water**

Wine/Enology-Grape Chemistry Group

